

1 **Length polymorphism at the *avpr1a* locus is correlated with male reproductive behavior in**
2 **a natural population of prairie voles (*Microtus ochrogaster*)**

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15 **Abstract** Laboratory studies have shown that vasopressin can influence sociosexual behavior
16 through its action on the vasopressin 1a receptor (V1aR). There is substantial evidence that the
17 length of a microsatellite in the gene (*avpr1a*) encoding for the V1aR can affect social
18 attachment to females and paternal behavior in male prairie voles under laboratory conditions.
19 However, previous field studies of prairie voles have failed to detect a strong effect of the length

20 of a male's *avpr1a* allele on their sociosexual behavior but these studies are typically much
21 shorter than the average prairie vole breeding lifespan. We examined the relationship between
22 male *avpr1a* microsatellite allele length and sociosexual behavior in a natural population of
23 prairie voles for 15 weeks, closer to the lifespan of prairie voles in nature. Contrary to
24 predictions, we found that males with the longest *avpr1a* microsatellite alleles were significantly
25 more likely to sire offspring with more than one female and to sire offspring that survived until
26 trappable age than males with the shortest *avpr1a* microsatellite allele lengths. This relationship
27 was the strongest for males with the longest tenure on the study site. As in previous field studies,
28 we did not find evidence of a relationship between a male's *avpr1a* genotype and any index of
29 social behavior including male residency status or the number of females with which males
30 associate. This is the first study to support the hypothesis that a male's *avpr1a* genotype is a
31 factor underlying variation in the genetic mating system of prairie voles under natural conditions.

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33 **Keywords** *Avpr1a* • Prairie vole • Reproductive success • Sociosexual behavior • Vasopressin

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40 **Introduction**

41 In many species of animals, individuals within as well as between geographically distinct
42 populations exhibit intraspecific variation in behaviors associated with reproduction and parental
43 care (e.g., Gross 1984, 1996; Brockmann 2008; Taborsky 2008; Wolff 2008; Schradin et al.
44 2012). Since variability in reproductive behavior can ultimately affect an individual's lifetime
45 reproductive success (Mock and Fujioka 1990; Gross 1996; Gubernick and Teferi 2000;
46 Schradin and Lindholm 2011), determining the mechanisms underlying this variation is crucial
47 for understanding the evolution of mating systems and social behavior. Empirical data from
48 numerous field studies suggest that intraspecific variation in reproductive behavior is frequently
49 an adaptive response to differences in environmental conditions (e.g., density; habitat structure;
50 Penteriani et al. 2011; Schradin et al. 2012) or individual characteristics (e.g., body size, age,
51 developmental stage, Young et al. 2007; Oliveira et al. 2008). More recently, an increasing
52 number of laboratory studies have provided an abundance of information on neurobiological
53 mechanisms that mediate sociosexual behavior both within as well as among species (Insel 1997;
54 Lim et al. 2004; Donaldson et al. 2010; McGraw and Young 2010). These studies suggest that
55 the basis for intraspecific variation in sociosexual behavior appears to be complex, perhaps
56 involving multiple ecological and neurogenetic influences.

57 Several neuropeptides can influence the expression of sociosexual behaviors in diverse
58 animal taxa (Witt 1995; Insel and Young 2000; Heinrichs and Domes 2008; Goodson and
59 Thompson 2010; Oldfield and Hofmann 2011; Godwin and Thompson 2012). In mammals,
60 behaviors such as social attachment, partner preference, and paternal care are mediated by the
61 neuropeptide vasopressin through its action on the vasopressin 1a receptor (V1aR; Hammock
62 and Young 2002; Hammock et al. 2005; Donaldson et al. 2010). Several lines of evidence

63 indicate that differences in the neural expression of V1aR contribute to inter- and intraspecific
64 variation in mammalian sociosexual behavior. Among arvicoline rodents, the socially
65 monogamous prairie voles (*Microtus ochrogaster*) and woodland voles (*M. pinetorum*) share
66 similar neural patterns of V1aR distribution and densities that differ significantly from those in
67 the nonmonogamous meadow (*M. pennsylvanicus*) and montane (*M. montanus*) voles (Hammock
68 and Young 2002; but see Fink et al. 2006). Male transgenic house mice (*Mus musculus*) and
69 meadow voles possessing the *avpr1a* gene from prairie voles have a neural distribution of V1aRs
70 similar to that in prairie voles and exhibit increased partner preference and affiliative behavior
71 towards females relative to nontransgenic males (Young et al. 1999; Lim et al. 2004). V1aR
72 signaling is necessary for the formation and expression of female partner preference in
73 experimentally manipulated male prairie voles. Experiments utilizing a viral vector to increase
74 the density of the V1aR in the ventral pallidum of male prairie voles increased partner preference
75 relative to control males (Pitkow et al. 2001). The opposite was found when RNA interference
76 was used to decrease expression of V1aR in the ventral pallidum of male prairie voles (Barrett et
77 al. 2013). These and other studies suggest that differences in the neural distribution and density
78 of V1aR can lead to variation in behaviors such as the formation of social attachments and
79 parental behavior in some mammals.

80 Although prairie voles are considered to be socially monogamous based on extensive
81 laboratory and field data (e.g., Carter et al. 1986; Getz et al. 1993), variation in social and genetic
82 monogamy occurs within all natural populations studied (Illinois: Getz et al. 1993; Kansas and
83 Indiana: Mabry et al. 2011; Streatfeild et al. 2011) as well as within populations maintained in
84 semi-natural enclosures (Ophir et al. 2008a; Solomon et al. 2009). In natural and semi-natural
85 populations, most male prairie voles are territorial (resident males) and reside at a nest with one

86 female but a sizeable minority (32 – 46 %) are nonterritorial (wandering males) with home
87 ranges that overlap multiple females (Getz et al. 1993; Solomon and Jacquot 2002; Ophir et al.
88 2008a). The degree of social monogamy varies within and between populations over time (Fitch
89 1957; Roberts et al. 1998; Cushing and Kramer 2005; Streatfeild et al. 2011). In addition, in
90 natural and semi-natural populations some males sire litters with multiple females while other
91 males sire offspring with just a single female (Ophir et al. 2008a; Ganey et al. 2009; Solomon et
92 al. 2009; Mabry et al. 2011). Multiple paternity has also been reported in litters from females in
93 natural (Illinois: Solomon et al. 2004; Indiana and Kansas: Mabry et al. 2011) and semi-natural
94 populations (Ophir et al. 2008a; Ganey et al. 2009; Solomon et al. 2009).

95 Some of the inter- and intrapopulation variation in sociosexual behavior observed in
96 prairie voles appears to be due to ecological factors such as population density and the
97 distribution of vegetation (Streatfeild et al. 2011) or differences in individual condition such as
98 body mass and parasite load (Chesh et al. 2012). There is also evidence that suggests that
99 individual differences in the pattern of V1aR neural expression may explain some of the
100 variability in reproductive behavior within or among populations. In prairie voles,
101 polymorphism in the length of the microsatellite DNA within the regulatory region of the gene
102 (*avpr1a*) encoding for V1aR predicts individual differences among males in the expression of
103 V1aRs in specific areas of the brain that appear to influence behaviors such as partner preference
104 and paternal care (Hammock and Young 2002, 2005; Hammock et al. 2005). Interestingly, there
105 were some inconsistencies among these studies with respect to which brain areas showed
106 differential V1aR expression, suggesting factors other than microsatellite length influence
107 expression. The differences in neural expression of V1aR in some brain regions are correlated
108 with some male sociosexual behavior under laboratory conditions (Pitkow et al. 2001; Hammock

109 and Young 2005). Specifically, in laboratory trials, males with longer *avpr1a* microsatellite
110 alleles exhibited greater expression of some behaviors characteristic of social monogamy
111 (partner preference, paternal care, affiliation) relative to males with shorter *avpr1a* alleles
112 (Hammock and Young 2005). These data suggest that a male's *avpr1a* genotype influences his
113 neural expression of V1aRs, which in turn affects his social behavior and provide a neurogenetic
114 mechanism for explaining some of the variation in male sociosexual behavior observed in natural
115 populations.

116 However, attempts to relate variation in male sociosexual behavior in field populations to
117 polymorphism in the length of a male's *avpr1a* microsatellite alleles have failed to show strong
118 support for this hypothesis. No significant relationships were detected between male *avpr1a*
119 genotype and indices of social and genetic monogamy from 4-8 week studies of natural
120 populations of prairie voles (Mabry et al. 2011). There was also no evidence that the *avpr1a*
121 genotype of male prairie voles predicted variation in several indices of social monogamy such as
122 space use or the likelihood of a male adopting a resident versus wanderer (male found at >1 nest)
123 tactic when semi-natural populations were studied for 3-weeks by Ophir et al. (2008b) or 15-
124 weeks by Solomon et al. (2009). In the Solomon et al. (2009) study, genetic parentage data
125 indicated that males with shorter *avpr1a* microsatellite alleles sired significantly more offspring
126 with more females relative to males with longer *avpr1a* microsatellite alleles, but no such
127 relationship was detected in the shorter-term Ophir et al. (2008b) study. Since laboratory
128 settings are much less complex than the field environment, it is not unusual to find that results
129 from a laboratory setting do not translate to the field (Calisi and Bentley 2009).

130 Recent work by Donaldson and Young (2013) provides direct evidence that the prairie
131 vole *avpr1a* microsatellite region influences neural V1aR expression in specific brain regions but

132 they also report that other genetic polymorphisms mediate neural V1aR expression and this may
133 be contributing to inconsistencies among studies. It has also been suggested that no simple
134 relationship exists between *avpr1a* allele length and behavior because length may be a rough
135 indicator of sequence differences that are the basis of the V1aR expression variation (Phelps et
136 al. 2010). Another reason for the inconsistencies in the studies in semi-natural and free-ranging
137 populations with respect to the relationship between male *avpr1a* genotype and reproductive
138 success may be the differences in length of the studies. The minimum interbirth interval among
139 female prairie voles is about three weeks (Keller 1985) and in the two shorter studies (Ophir et
140 al. 2008b; Mabry et al. 2011) almost all females had just a single litter while in the study by
141 Solomon et al. (2009) females had up to four litters. Since the lifespan of prairie voles averages
142 30-122 days depending on factors such as density, season of birth and dispersal (Getz et al.
143 1994), a longer time interval may need to be studied to more accurately assess the relationship
144 between *avpr1a* polymorphism and sociosexual behavior in free ranging populations,
145 particularly if the *avpr1a* microsatellite only has a modest influence on neural V1aR expression
146 as suggested by Donaldson and Young (2013).

147 Therefore, the objective of this study is to examine the relationship between male *avpr1a*
148 genotype and sociosexual behavior in a free ranging population of prairie voles over a period of
149 time similar to the average adult lifespan to test the hypothesis that *avpr1a* length polymorphism
150 is contributing to the variation in male social attachment and mate fidelity observed in free-living
151 populations. If microsatellite length polymorphism at the *avpr1a* locus is an important factor
152 contributing to individual differences in sociosexual behavior in nature, we expected to detect
153 relationships between male *avpr1a* genotype and some indices of social and reproductive
154 behavior measured in the field. Specifically, as suggested by the partner preference study by

155 Hammock and Young (2005) and supported by the results of our study in semi-natural
156 populations (Solomon et al. 2009), we expected that males with longer *avpr1a* microsatellite
157 alleles would have a higher residency score (i.e., be classified as residents more often than as
158 wanderers), associate with a smaller proportion of females, show a greater amount of home range
159 overlap with the female that the male overlapped with most, associate with her the most, and sire
160 offspring with fewer females relative to males with shorter *avpr1a* microsatellite alleles. If the
161 number of females that a male sires offspring with is closely associated with his reproductive
162 success then males with longer *avpr1a* microsatellite alleles should have lower reproductive
163 success (i.e., sire fewer offspring) than males with shorter *avpr1a* microsatellite alleles.

164

165 **Materials and methods**

166 Study site and animals

167 The study site was located approximately 5 km north of Bloomington, Indiana, within the
168 Indiana University Bayles Road Preserve (39°13'00"N, 86°32'27"W). The site is an old field,
169 primarily composed of grasses and forbs and is maintained by periodic mowing to prevent
170 successional changes. Data were collected from a 10,000 m² (1 ha) area during a 15-week period
171 from early-June to mid-September 2010, coinciding with the mid- to late-breeding season of
172 prairie voles (Getz et al. 1993). We monitored prairie voles within the study area by live-
173 trapping using Ugglan multiple-capture traps (Grahnb, Hillerstorp, Sweden). Traps were baited
174 with cracked corn, a low quality food item (Desy and Batzli 1989; Cochran and Solomon 2000)
175 and shielded from heat and rain by aluminum flashing covered with vegetation. Every time an
176 individual was captured, we recorded the following: location of capture, identification number,

177 sex, body mass, age class, reproductive status (males: scrotal or non-scrotal, females: pregnant
178 [determined by gently palpating a female's abdomen], lactating, or non-reproductive), body mass
179 and any individuals with which they were captured. Upon first capture, all individuals were
180 given a unique toe clip for identification and these tissue samples were stored at -20 °C for future
181 genetic analysis. Body mass was determined to the nearest gram using a Pesola micro-line
182 spring scale (Forestry Suppliers Inc., Jackson, MS, USA). Animals >29 g in mass were
183 classified as adults, while animals 21-29 g were considered subadults, and animals <21 g were
184 classified as juveniles (Gaines et al. 1979; Getz et al. 1993).

185 During the first three weeks of the study, we trapped on a 21x6 grid with grid points
186 approximately 10 m apart and one trap per grid point. During each grid-trapping week, we
187 checked the traps eight times. Traps were open from 2000 on Sunday until 0700 on Monday
188 when they were checked and left unset. Traps were reset at 1800 on Monday, Tuesday,
189 Wednesday, and Thursday nights with trap checks taking place on those nights at 2000 as well as
190 0700 the next mornings with the exception of Wednesday (i.e., Tuesday, Thursday and Friday
191 mornings). Traps were set from 1800 to 2000 and overnight until 0700 (except Wednesdays) to
192 avoid heat related trauma to trapped animals. At all other times, traps were left in place but
193 unset. Grid trapping allowed us to monitor population density, estimate home range size and
194 space use, and capture females for radio-tracking to their respective nests.

195 We radio collared all adult females caught during the first 3 weeks of grid trapping and
196 used radio-telemetry to locate their nests. Females to be radio tracked were fitted with model
197 PD-2C radio transmitters (Holohil Systems Ltd, Ontario, CA) attached around the neck of the
198 individual with a small zip tie covered in rubber tubing (see Keane et al. 2007; Lucia et al. 2008
199 for details). The radio collars weighed approximately 3 g and were always less than 10% of a

200 female's body mass. A radio collar of less than 10 % of a female's body mass should not
201 significantly influence her daily energy expenditure (Berteaux et al. 1996). Furthermore,
202 Pouliquen et al. (1990) found no negative effects on social interactions of wild mice, *Mus*
203 *domesticus*, after attachment of radio collars. Radio-tracking was conducted via triangulation
204 using two FieldMaster receivers (Johnson's Telemetry, Ed Dorado Springs, MO) and three-
205 element Yagi antennas (Johnson's Telemetry, Ed Dorado Springs, MO). Fixes were taken for
206 each female twice a day, once between 0900 and 1000 and again between 1600 and 1700,
207 because it was assumed that females would likely be underground at these times. Nest locations
208 were determined by searching for surface nests or entrances to underground nests in the vicinity
209 where females were tracked. A female had to be tracked to the same nest location for a
210 minimum of 3 consecutive days to confirm that she was living at that nest location. Once a nest
211 location was confirmed, we placed three Ugglan traps in the surface runways within 1 m of nest
212 entrances and the female's radio collar was removed during her next capture. If a female's nest
213 was not determined by the end of the 5th week, the collar was removed upon the female's next
214 capture.

215 After the third week of grid trapping, we switched to nest trapping for two weeks,
216 followed by one week of grid trapping. During the last 12 weeks of the study, we divided the
217 trapping into 4 periods, with each period composed of two nest trapping weeks and the following
218 grid trapping week. During nest trapping weeks, only the traps at the female nests were checked.
219 We checked nest traps 10 times during each nest trapping week. Nest trapping followed the
220 same schedule as grid trapping but we also set traps at 1800 on Sundays and Fridays followed by
221 checks at 2000 on those days. As with grid trapping, traps were left in place but unset at all other

222 times. Nest trapping allowed us to determine which individuals were residents at nests, which
223 adult males associated with each adult female, and to capture juveniles for parentage analysis.

224 Adult prairie voles may change residency status (resident versus wanderer) or social
225 associations with females during their lifetime (Solomon and Jacquot 2002; McGuire and Getz
226 2010). Therefore, we determined adult male space use, residency status, and social associations
227 with adult females separately for each of the 4 trapping periods during the final 12 weeks of the
228 study.

229

230 Space use

231 Traditional estimators of home range size were not useful for this study because many voles
232 were captured only a few times. The number of captures per individual ranged from 1 to 109,
233 with 47.5% of voles being captured ≤ 5 times. When individuals are trapped only a few times,
234 the mean square distance (MSD) moved by individuals from their center of activity can be a
235 suitable method to estimate home range size, because only 3 different capture locations are
236 needed to calculate the geometric center of a two-dimensional region (e.g., home range, Slade
237 and Russell 1998). Slade and Russell (1998) showed that the MSD moved by prairie voles from
238 their center of activity was more highly correlated with other estimates of home range size, such
239 as minimum convex polygon, harmonic mean or kernel densities, than other measurements of
240 distance moved. Therefore, we estimated adult male and female prairie vole home range size by
241 calculating the MSD moved from an individual's center of activity (Slade and Swihart 1983;
242 Diffendorfer et al. 1995; Streatfeild et al. 2011). A vole's center of activity was calculated as the
243 mean X and Y coordinates of all an individual's captures from grid trapping and represents the

244 geometric center of an individual's grid capture locations (Hayne 1949). For each of the four 3-
245 week periods of trapping data, we calculated the MSD of all adults trapped at ≥ 3 different
246 locations during the grid trapping week of a particular trapping period. Home range sizes were
247 not calculated for the fourth time period, due to the presence of a raccoon on the field site that
248 disturbed many traps during the last week of grid trapping.

249 The number of females with which a male's home range overlapped was quantified from
250 grid trapping data for each 3-week trapping period. In addition, for each trapping period we
251 calculated the percentage overlap of a male's home range with the home range of the female that
252 the male's home range overlapped the most. A male and female were considered to overlap
253 (share a trap location) if each were captured at least once during the same grid-trapping week at
254 the same trap location but not necessarily the same time or day. To determine the proportion of
255 overlap with the most overlapped female, the number of shared grid trap locations with the
256 female with which the male shared the most trap locations was divided by the total number of
257 locations at which the male was trapped (Streatfeild et al. 2011).

258

259 Social associations and nest residency

260 To evaluate the number of female social ties per male, we determined the number of unique
261 females with which each male associated during each 3-week trapping period and the pairwise
262 half-weight association index (AI; Cairns and Schwager 1987; Whitehead 1999, 2009; Mabry et
263 al. 2011) between every male and each female with which he associated during each period. The
264 AIs were calculated using SOCPROG software and customized coding in R. A male was
265 defined as "associating" with a female when both were captured at the same nest, but not

266 necessarily in the same trap, during the same trap check. We also determined a male's relative
267 association index (relative AI) with the female with which he associated most, to quantify the
268 relative strength of each male's social interactions with the female(s) with which he associated.
269 Relative association index was calculated by dividing a male's AI with the 'most associated
270 female' by the sum of the AIs with all females with which he associated during a given trapping
271 period (Mabry et al. 2011). Relative AI is conceptually very similar to the relative encounter rate
272 used by Ophir et al. (2008a) to quantify the relative overlap of male and female prairie vole
273 home ranges. A relative AI of ≥ 0.5 indicates that a male associated with a given female more
274 often than with all other females combined and a relative AI value of 1 indicates that a male
275 associated with only one female.

276 Data from nest trapping weeks was used to determine the residency status of sub-adult
277 and adult males. Since males may change their residency status during their lifetime (Solomon
278 et al. 2009; McGuire and Getz 2010), we determined the residency status of males for each 3-
279 week trapping period. Sub-adult and adult males were classified as a resident at a particular nest
280 if $\geq 75\%$ of their captures were at one nest during a particular 3-week trapping period and they
281 were captured at least once during each nest trapping week during that period (Cochran and
282 Solomon 2000). Conversely, sub-adult and adult males that did not have $\geq 75\%$ of their captures
283 at a single nest site but were captured at least once during each nest trapping week during a 3-
284 week time period were classified as wanderers. Males not captured at least once during each
285 nest-trapping week within a particular 3-week trapping period were considered likely to be
286 visitors that lived outside the trapping grid and were not included in any analyses. For each
287 trapping period, residents were given a residency score of one while wanderers received a score
288 of zero. A male's average residency score over the four trapping periods was used as an index of

289 the extent to which a male adopted a resident versus a wanderer strategy (Solomon et al. 2009).
290 Trapping periods when a male could not be classified as a resident or wanderer were excluded
291 from the calculation of a male's residency score.

292

293 *Avpr1a* microsatellite analysis

294 We determined the *avpr1a* genotypes of males using the polymerase chain reaction (PCR) to
295 amplify the *avpr1a* microsatellite region of the V1aR using primers designed specifically for
296 prairie voles (Hammock and Young 2005; Solomon et al. 2009; Mabry et al. 2011). Genomic
297 DNA for microsatellite analysis was extracted from toe clips using DNeasy extraction kits
298 (Qiagen, Valencia, CA, USA). PCR amplification was conducted in 15 μ l reactions containing
299 100 ng/ μ l DNA, 10 mM Tris-HCL, 0.2 mM dNTPs, 1.5 mM MgCl₂, 0.67 μ M of the forward
300 (fluorescently labeled with 6-FAM phosphoramidite; IDT DNA Technologies, Coralville, IA,
301 USA) and reverse primers and 0.5 U of GoTaq DNA polymerase in the supplied reaction buffer
302 (Promega Corp, Madison, WI, USA). After denaturation at 94 °C for 2 min, PCR reactions were
303 cycled 25 times, with denaturation at 94 °C for 15 s, annealing for 20 s at 52 °C and extension at
304 72 °C for 45 s followed by a final elongation for 7 min at 72 °C. PCR products were diluted and
305 combined with an internal size standard (ROX GS2500, Applied Biosystems, Foster City, CA,
306 USA) and fragments detected using an ABI 3730 DNA sequencer with 50 cm capillary length
307 (Applied Biosystems, Foster City, CA, USA). Base pair (bp) lengths of PCR products were
308 determined using Genemapper (version 3.7) fragment analysis software (Applied Biosystems,
309 Foster City, CA, USA). The *avpr1a* microsatellite region consists of multiple types of repeat
310 units that makes binning the fragments into discrete allele size categories difficult. Therefore,

311 we quantified the microsatellite allele lengths using the raw bp measurements from GeneMapper
312 (version 3.7) and treated *avpr1a* allele length as a continuous variable (Mabry et al. 2011).

313

314 Parentage analysis

315 Genomic DNA used for analyzing male *avpr1a* genotypes was also used for parentage analysis.

316 To assess parentage, all live-trapped voles were genotyped at six microsatellite loci shown to be

317 polymorphic in prairie voles using PCR to amplify microsatellite DNA (for details of PCR

318 reactions see Keane et al. 2007; Solomon et al. 2009). PCR products were diluted and combined

319 with an internal size standard (LIZ GS500, Applied Biosystems, Foster City, CA) and fragments

320 were detected using an ABI 3730 DNA sequencer with 50 cm capillary length (Applied

321 Biosystems, Foster City, CA, USA). Base-pair lengths of the fluorescently labeled DNA

322 fragments were analyzed using Genemapper (version 3.7) fragment analysis software (Applied

323 Biosystems, Foster City, CA, USA) and microsatellite alleles compiled into discreet size classes

324 using FlexiBin (Amos et al. 2006).

325 We used the program Cervus 3.0 to calculate the polymorphic information content, and

326 observed and expected heterozygosity at each locus used in the study. For each locus, Cervus

327 tests if genotypic distributions deviated from Hardy–Weinberg expectations using chi-square

328 goodness of fit tests (Kalinowski et al. 2007). Cervus 3.0 was also used to conduct a genetic

329 determination of parentage for juveniles trapped in the field (Kalinowski et al. 2007). The

330 Cervus 3.0 software calculates a likelihood score for each candidate parent to identify the male

331 and female that was most likely to be the biological parents of a specific offspring. The

332 statistical confidence of these parentage assignments is calculated using a simulation that takes

333 into account population allele frequencies, an estimate of genotyping error, proportion of missing
334 genotypes, total number of candidate parents sampled, and the proportion of candidate parents
335 sampled. The simulation was performed for 10,000 cycles with a genotyping error rate of 0.02.
336 This error rate was based on empirical estimates of two potential sources of error: mutation and
337 mis-scoring of alleles (Solomon et al. 2004). The remaining input parameters for the simulation
338 were based on the actual data from the study population.

339 For each juvenile trapped in the field, only individuals classified as adults or older sub-
340 adults (25-29 g) at least five days prior to the date a particular juvenile was estimated to have
341 been conceived were considered candidate parents for parentage analysis. The birth dates of
342 individuals first trapped as juveniles were estimated based on their body mass from live-trapping
343 data. Pups typically weighed 2-3 g at birth and gain approximately 0.6-1.0 g daily (unpubl. data)
344 until weaning (21 d). The conception date was then estimated by subtracting 21 d from the
345 estimated birth date since 21 d is the average length of gestation for prairie voles (Richmond and
346 Conway 1969; Nadeau 1985). We used this method to estimate the date of conception for each
347 juvenile to determine candidate parents for parentage analysis. For each juvenile, we only
348 considered candidate parents trapped within 20 m of the juvenile's site of first capture (either a
349 natal nest or grid trap) as possible biological parents (see Winters and Waser 2003; Mabry et al.
350 2011). A distance of 20 m was used in the analysis because it is the approximate average home
351 range diameter of adult prairie voles in this population (Streatfeild et al. 2011).

352 Parentage was assigned using the parent-pair analysis option in Cervus 3.0 and a male
353 and female were accepted as the parents of a particular juvenile only if Cervus assigned both
354 parents with a confidence level of $\geq 95\%$. If after the initial analysis, a mother but not a father
355 was assigned as a parent with at least 95 % confidence, we reran the analysis using the 'known

356 mother' option in Cervus 3.0, again only considering adult males captured within 20 m of the
357 juvenile's site of origin as candidate fathers. We used the genetic parentage data to determine
358 the total number of offspring sired by each male and the number of female partners with which a
359 male sired offspring. We could only determine how many different females a male successfully
360 sired young with, not necessarily all of the females with which a male mated.

361

362 Ethical note

363 All procedures involving the trapping, marking and handling of prairie voles were
364 approved by the animal care and use committees of Miami University and Indiana University,
365 and were consistent with the guidelines published by the American Society of Mammalogists
366 (ASM; Sikes et al. 2011) for the use of wild animals in research.

367 We individually marked prairie voles in the field using uniquely-coded combinations of
368 toe clips. Although more benign marking methods are available for marking rodents, we believe
369 that none of them were as suitable for permanently marking field populations of prairie voles as
370 toe-clipping. Non-toxic dyes and hair clipping can be used to mark mammals but these
371 identifying marks are temporary because the fur is shed or re-grows (Johnson 2001; Sikes et al.
372 2011). Since we studied the field population for almost four months, we needed to have a means
373 of marking the animals with a more permanent identification mark. Also, these techniques are
374 not appropriate for marking altricial neonates that are hairless. More permanent marking
375 methods for field populations of mammals recommended by the ASM (Sikes et al. 2011) include
376 freeze branding, ear-notching, tattooing, ear tagging, passive integrated transponders (PIT tags)
377 and toe-clipping. Each of these procedures probably causes some amount of stress or pain to the

378 animal. Freeze-brands, ear notches and tattoos were inappropriate for this study because of the
379 limited number of unique marks that can be made, particularly on newly caught neonates. Adult
380 prairie voles have very small external ear pinnae and ear tag loss in prairie voles has been
381 estimated to be 10-16% (Wood and Slade 1990; Harper and Batzli 1996) when the tags were lost
382 by being ripped from the ears (e.g., when caught on vegetation) of tagged individuals. In
383 addition to causing pain, the loss of ear tags could compromise the integrity of our trapping data
384 since individuals cannot be positively identified after the loss of their tags. Finally, Wood and
385 Slade (1990) found no detrimental effects of toe-clipping on prairie vole survival and body mass
386 relative to ear tagging. Recent advances in technology have made it possible to mark animals for
387 individual identification using PIT tags implanted subcutaneously. We believe this method has
388 several drawbacks for marking prairie voles relative to toe-clipping. First, the small size of
389 juveniles may preclude the use of PIT tags. There is no published information that indicates how
390 large an animal must be to safely inject a PIT tag. Injecting a PIT tag into a juvenile prairie vole
391 may be risky due to the invasive nature of the procedure and the gauge of the needle needed to
392 inject the tag. Second, the invasive nature of the procedure also leaves implant wounds that can
393 become infected in voles (Harper and Batzli 1996). A study by Harper and Batzli (1996) also
394 found that 5% of the voles PIT tagged lost their tag which could compromise our ability to track
395 specific individuals through time.

396 The ASM permits the use of toe-clipping to mark animals when none of their other
397 recommended marking methods appear suitable, especially if tissue samples also need to be
398 collected (Sikes et al. 2011). We felt that toe-clipping was the most feasible method of
399 permanently marking the large numbers of prairie voles of varying size classes in our study and
400 it also allowed us to collect tissue samples for parentage analysis. We used a clean, sharp pair of

401 scissors to remove toes and no more than 1 toe per foot was clipped. The ASM guidelines also
402 do not recommend the use of anesthetics or analgesics during toe-clipping because of the
403 prolonged period of restraint that is necessary to apply them, and because consumption of
404 analgesic substances by licking may cause additional stress to the animal (Sikes et al. 2011).
405 Toes were stored in 1.5 ml microcentrifuge tubes at -20 °C until DNA was extracted for
406 parentage analysis.

407

408 Statistical analysis of data

409 Previous studies have demonstrated that analyses of the effect of male *avpr1a* microsatellite
410 allele length on social and reproductive behavior yield similar results when using the summed
411 length of a male's two *avpr1a* microsatellite alleles or either just the longer or shorter of a male's
412 two *avpr1a* microsatellite alleles as the independent variable (Hammock et al. 2005; Ophir et al.
413 2008b; Solomon et al. 2009; Mabry et al. 2011). Therefore, we examined the correlation
414 between male *avpr1a* genotype and all indices of social and reproductive behavior using the sum
415 of a male's two *avpr1a* microsatellite lengths as the metric of a male's *avpr1a* genotype (additive
416 genetic model). We determined if the distribution of summed *avpr1a* microsatellite allele
417 lengths among males differed from a normal distribution with a Shapiro-Wilk test.

418 We analyzed the indices of space use and social associations during each of the four 3-
419 week trapping periods separately, so that we could monitor changes in an individual male's
420 behavior (e.g., residency status, home range size, overlap with females and association indices),
421 throughout the last 12 weeks of the study. The relationship between male *avpr1a* microsatellite
422 length and the mean square distance moved by males or the proportion of females in the

423 population overlapped by males was analyzed using linear regression with male *avpr1a*
424 microsatellite length as the independent variable. Due to the uneven distribution of the response
425 variable, the proportion of home range overlap between males and the adult female each male
426 overlapped with the most was examined using logistic regression, with male *avpr1a*
427 microsatellite length as the independent variable. A value of 1 was assigned to males whose
428 proportional overlap with their most overlapped female was ≥ 0.75 , while male's whose
429 proportional overlap with their most overlapped female was < 0.75 were assigned a value of zero.
430 A value of 0.75 was used for categorizing a male's overlap with his most overlapped female
431 since males and females that had 75 % of their captures at the same nest site were considered to
432 be co-residents.

433 The relationships of male *avpr1a* microsatellite length with the number of females with
434 which a male associated (AI) and the relative strength of a male's association with the most
435 associated female (relative AI) were examined using logistic regression with male *avpr1a*
436 microsatellite length as the independent variable. We choose to use a logistic regression model
437 because there were not enough different response values to make a continuous-type analysis
438 reasonable since most males associated with 1 or 2 females. If a male associated with only 1
439 female, an AI value of 0 was assigned, whereas if a male associated with more than 1 female an
440 AI value of 1 was assigned for analysis. Relative AI values < 1 were given a value of 0 and
441 relative AI values of 1 remained at a value of 1 for analysis. These values were assigned for both
442 AI and relative AI to compare socially monogamous males (associating with only one female)
443 and non-socially monogamous males (associating with > 1 female) against their respective
444 *avpr1a* genotype. Additionally, the relationship of male *avpr1a* microsatellite length with a
445 male's average residency score over the four trapping periods was examined using logistic

446 regression, where a value of 0 was assigned to all voles with an average score <1 (wanderers)
447 and a value of 1 assigned to all voles with an average score equal to 1 (residents).

448 While measures of most social indices were evaluated separately during each of the four
449 3-week trapping periods in order to account for changes in social behavior through a male's
450 lifespan, the number of offspring sired per male and number of females with which a male sired
451 offspring were calculated over the entire length of the study to estimate lifetime reproductive
452 success and genetic monogamy. The relationship between *avpr1a* microsatellite length and
453 whether or not a male sired any offspring were conducted using logistic regression. Males that
454 sired any offspring were assigned a value of 1, whereas a value of 0 was assigned to males that
455 did not sire any offspring.

456 The number of offspring sired per male and the number of females with which a male
457 sired offspring were ordinal values that fell within a narrow range. Therefore, the relationship
458 between male *avpr1a* microsatellite lengths with these two dependent variables was analyzed
459 using ordinal regression, since the number of offspring and female partners can easily be
460 partitioned into several ordinal categories. If a male did not sire any offspring, a value of 0 was
461 assigned. A value of 1 was assigned to males that sired 1 offspring and a value of 2 was
462 assigned to males that sired >1 offspring. For the ordinal regression that examined the
463 relationship between male *avpr1a* microsatellite length and the number of reproductive partners,
464 males that did not have any reproductive partners were assigned a value of 0, a value of 1 was
465 assigned to males with 1 reproductive partner and a value of 2 was assigned to males with >1
466 reproductive partner. In addition, the length of time adult males were known to be in the study
467 population (an index of survival) was included as a covariate in the ordinal regression because
468 males that were present on the study grid for just a few weeks likely sired fewer offspring with

469 fewer females irrespective of male *avpr1a* genotype relative to males that were present most of
470 the study. The length of time adult males were known to be in the study population (hereafter
471 referred to as tenure) was measured as the number of days between a male's first and last
472 capture.

473 The ordinal regression model used here is based upon the cumulative logit function. That
474 is, it models a series of log odds by way of a linear combination of predictors. More specifically,
475 and simplified to include just a single continuous predictor for clarity,

$$476 \quad \text{logit } P(Y \leq j) = \log \left[\frac{P(Y \leq j)}{1 - P(Y \leq j)} \right] = \alpha_j + \beta_1 X_1$$

477 where Y is the response (e.g., number of partners), j represents response category j (e.g. 0, 1, or
478 more than 1 partner), α_j is an intercept unique to response category j , and X_1 is some
479 continuous predictor. Inherent in this model is the assumption that the same regression
480 coefficients (β_1 in the example above) can be used to represent the relationship between the
481 predictor and the logit, for any response category j . Therefore, in the so-called proportional odds
482 model, there is a single set of coefficients in the model (Hilbe 2009). The proportional odds
483 model was fit using the VGAM package in R (Yee 2010). We tested this assumption for the
484 models presented in this paper using a likelihood ratio test (Yee 2010), and found that the
485 proportional odds assumption was appropriate.

486 In order to use an ordinal regression to analyze the number of female mates and number
487 of offspring against summed *avpr1a* microsatellite length, both male *avpr1a* microsatellite length
488 and tenure were categorized into quartiles (*avpr1a* [bp length]: Q1 = 1390.34-1456.59, Q2 =
489 1456.6-1469.45, Q3 = 1469.64-1481.29, Q4 = 1481.3-1523.22; tenure [d]: Q1 = 1-21, Q2 = 22-
490 40, Q3 = 41-73, Q4 = 74-97), where *avpr1a* microsatellite length was the independent variable

491 and tenure was a covariate. This was done because we were unable to check the aforementioned
492 proportional odds assumption with the predictors as continuous variables. Instead, as is typical
493 in modeling categorical predictors, we made the categorized version of each predictor (*avpr1a*
494 and tenure) into three dummy variables.

495 All statistical analyses were conducted using statistical software R version 2.13.2. The
496 average number of offspring sired by each male and the average number of female partners with
497 which each male sired offspring are reported as mean \pm 1 SE. A $P\leq 0.05$ was considered
498 statistically significant for all analyses.

499

500 **Results**

501 *Avpr1a* genotyping

502 Of the 238 adult and subadult male prairie voles that were caught on the study site, we were able
503 to genotype 168 (70.6 %) of these males at their *avpr1a* microsatellite locus. Summed *avpr1a*
504 lengths ranged from 1390 to 1570 bp with a median length of 1470 bp. The distribution of
505 summed *avpr1a* lengths was statistically different from a normal distribution with proportionally
506 more males with summed *avpr1a* lengths of 1470-1510 bp in the population than expected in a
507 normal distribution (Shapiro-Wilk test: $W=0.97$, $N=168$, $P=0.0006$, Fig. 1).

508

509 Social behavior

510 The results of the analyses of social behavior are shown in Table 1. Male *avpr1a* microsatellite
511 allele length did not significantly predict any of the indices of adult male space use or social

512 associations with females during any trapping period. Home range sizes (MSD) of adult males
513 did not differ with male *avpr1a* microsatellite allele length. Adult male *avpr1a* microsatellite
514 allele length did not predict either the proportion of adult females overlapped by males or the
515 proportion of the home range overlap with the adult female a male overlapped with the most.
516 Neither the number of adult females with which an adult male associated (AI) nor the relative
517 strength of a male's association with his most associated female (Relative AI) was predicted by
518 male *avpr1a* microsatellite allele length. Finally, male *avpr1a* microsatellite allele length did not
519 predict an adult male's average residency score.

520

521 Genetic parentage assignment and reproductive success

522 The summary statistics for the six microsatellite loci used in the parentage analysis are shown in
523 Table 2. We detected relatively high levels of polymorphism across loci, with 18-28 alleles per
524 locus. A single locus (MSMM-5) exhibited a significant departure from Hardy-Weinberg
525 equilibrium, likely due to the presence of null alleles as indicted by Cervus 3.0. However, since
526 the frequency of null alleles at this locus was not likely to substantially bias exclusion
527 probabilities for parentage analysis (Dakin and Avise 2004), we utilized all six loci in assigning
528 parentage. The polymorphic information content of the loci ranged from 0.428 to 0.902.

529 Overall, we were able to assign parent-pairs with a 95 % confidence level for 82 of 139
530 (59 %) individuals first caught as juveniles, which is not uncommon in open populations of
531 prairie voles in nature (Mabry et al. 2011). Ten of the juveniles that we were able to assign both
532 parents using Cervus 3.0 were first trapped with an adult female ($N=10$ females) from which they
533 were nursing. In all 10 cases the female that was assigned as the biological mother of the

534 juvenile using Cervus 3.0 was also the female that the juvenile was nursing from when first
535 trapped, providing an independent corroboration of maternity for these juveniles.

536 Of the 144 males that were considered as candidate fathers for the parentage analysis,
537 only 38 (26 %) of these males sired the 82 juveniles for which we could assign both a male and
538 female parent. These 38 males that sired offspring had an average of 1.5 ± 0.9 female mates
539 (range 1-5). We found evidence that male *avpr1a* microsatellite allele length was associated
540 with the number of females with which males sired offspring ($N=144$; $t=-1.54$, $P=0.02$). This
541 implies that the probability of siring offspring with no females was greater among males with the
542 shortest *avpr1a* lengths (those in the 1st quartile) than males with the longest *avpr1a* lengths
543 (those in the 4th quartile) across all tenure categories (Table 3). Furthermore, males with the
544 longest *avpr1a* lengths (those in the 4th quartile) were more likely to sire offspring with 1 female
545 or more than 1 female relative to males with the shortest *avpr1a* lengths (those in the 1st quartile)
546 in every tenure category (Fig. 2).

547 The 38 males that successfully sired offspring had an average of 1.7 ± 1.2 offspring (range
548 1-6), with 23 males (61 %) siring only 1 offspring. Overall, we did not find evidence that *avpr1a*
549 microsatellite length predicted whether or not a male sired offspring ($N=144$; $z=1.57$, $P=0.12$).
550 However, evidence existed that *avpr1a* microsatellite length was associated with the number of
551 offspring produced by males ($N=144$; $t=-1.54$, $P=0.02$). Again, this implies that the probability
552 of having zero offspring decreased between males from the 1st quartile to those in the 4th quartile
553 across all tenure categories (Fig. 3), meaning that males with the longest *avpr1a* lengths (those in
554 the 4th quartile) were less likely to have zero offspring than males with the shortest *avpr1a*
555 lengths (those in the 1st quartile). Across all tenure categories, males with the longest *avpr1a*

556 lengths (those in the 4th quartile) were more likely to have 1 offspring or more than 1 offspring
557 than males with the shortest *avpr1a* lengths (those in the 1st quartile; Fig 3).

558

559 **Discussion**

560 Although previous studies have demonstrated a relationship between a male prairie vole's
561 *avpr1a* genotype and his sociosexual behavior in laboratory (Hammock and Young 2005) and
562 semi-natural settings (Solomon et al. 2009), this is the first study to document a relationship
563 between a male's *avpr1a* genotype and reproductive behavior in a natural population of prairie
564 voles. We found that a male's *avpr1a* genotype predicted whether a male sired offspring with
565 zero, one or more than one female as well as the number of offspring sired. Contrary to our
566 predictions, males with the longest *avpr1a* microsatellite alleles were significantly more likely to
567 sire offspring with more than one female and to sire one or more offspring than males with the
568 shortest *avpr1a* microsatellite allele lengths. The relationship between a male prairie vole's
569 *avpr1a* genotype and his reproductive behavior was most evident among adult males with the
570 most extreme differences in *avpr1a* allele length. For example, when the reproductive data was
571 analyzed using ordinal regression after categorizing *avpr1a* and tenure into just two groups, i.e.,
572 below and above the median, instead of into quartiles, there was not an apparent effect of *avpr1a*
573 on the number of females with which a male sired offspring or his reproductive success.
574 Furthermore, the relationship between a male's *avpr1a* genotype and his reproductive behavior
575 was strongest among adult males with the longest tenures (> 40 d) on the study site. This
576 suggests that the impact of male's *avpr1a* genotype on his reproductive behavior appears
577 relatively weak, as suggested by Donaldson and Young (2013), but can result in significant
578 differences among males when reproductive behavior is measured over a time frame similar to

579 the average lifespan of prairie voles, illustrating the importance of long-term field studies.
580 Finally, our findings support the hypothesis that one factor responsible for the inconsistencies
581 among the previous field studies examining the relationship between male *avpr1a* genotype and
582 reproductive behavior was variability in the length of the studies.

583 Although the laboratory studies suggest that males with longer *avpr1a* microsatellite
584 alleles should be more socially monogamous in nature since they spend more time in contact
585 with their female partner and display less interest in unfamiliar females (Hammock and Young
586 2005), we failed to detect correlations between a male's *avpr1a* genotype and residency status,
587 space use or social associations with females. These findings are consistent with all of the
588 previous field studies of prairie voles that have also failed to detect any relationship between
589 male *avpr1a* genotype and indices of male space use or social association with females (Ophir et
590 al. 2008b; Solomon et al. 2009; Mabry et al. 2011). The field investigations of the relationship
591 between male *avpr1a* genotype and measures of male space use and social association with
592 females indicate that, at least for the indices that have been examined, if a male's *avpr1a*
593 genotype influences these behaviors in nature the effects are weak, relative to other genetic and
594 environmental factors that mediate these behaviors (Ophir et al. 2008b; Solomon et al. 2009;
595 Mabry et al. 2011). Perhaps indices of male social behavior with a more fine scale resolution
596 may reveal stronger correlations. Our findings regarding social behavior also suggest that the
597 greater reproductive success of males with the longest *avpr1a* alleles was not due to differences
598 in residency status, space use or social associations with females among males with different
599 *avpr1a* allele lengths.

600 Males with shorter *avpr1a* alleles spend more time investigating unfamiliar females than
601 males with longer *avpr1a* alleles in laboratory trials (Hammock and Young 2005). Therefore, we

602 predicted that males with shorter *avpr1a* alleles would mate with more females and engage in
603 more extra-pair copulations (EPCs) in natural populations, resulting in these males siring
604 offspring with more females and perhaps more total offspring. Contrary to our expectations, it
605 was males with longer *avpr1a* alleles that produced offspring with more females and sired more
606 offspring. In our study, we could not assess the number of matings per male but only the number
607 of offspring sired per male that survived to trappable age. One reason that males with longer
608 *avpr1a* alleles produced offspring with more females and sired more offspring may have been
609 because the offspring of these males were more likely to survive to be trapped as juveniles and
610 not because the males mated with more females. In laboratory studies, males with longer *avpr1a*
611 microsatellites spent more time with their female social partner and lick and groom pups more
612 than males with shorter *avpr1a* microsatellite lengths (Hammock and Young 2005). Since
613 prairie voles are socially monogamous (Carter and Getz 1993) and exhibit biparental care
614 (Solomon 1993; Lonstein and De Vries 1999), these differences in behavior may have
615 contributed to the greater reproductive success of males with the longest *avpr1a* alleles. In the
616 laboratory, removal of the male partner following mating significantly decreased the probability
617 that a female would give birth compared to females whose mates were present throughout the
618 entire pregnancy (McGuire et al. 1992; Dewsbury 1995). Laboratory-reared offspring also ate
619 solid food earlier when their father was present (Wang and Novak 1992). Although McGuire et
620 al. (1992) found no effect of male presence on the survival of prairie vole offspring maintained
621 in the laboratory, which is not surprising since food is always available and there is no predation
622 pressure, when laboratory-reared animals were released into field enclosures the presence of the
623 female's male social partner significantly increased offspring survival (Mahady and Wolff 2002).
624 Data from laboratory studies also suggest that males with longer *avpr1a* microsatellite alleles

625 may be more desirable as social partners because they would be likely to spend more time at the
626 nest and provide more paternal care than males with shorter *avpr1a* microsatellite alleles, thereby
627 increasing a female's reproductive success. Consistent with this hypothesis, Castelli et al. (2011)
628 showed that females in laboratory preference trials where they were given access to males with
629 long and short *avpr1a* alleles, spent more time with and mated more frequently with males with
630 longer *avpr1a* microsatellite alleles. Thus the greater number of females that males with longer
631 *avpr1a* alleles sire offspring with is not necessarily because these males engage in more EPCs.
632 Males with longer *avpr1a* alleles may be more likely to have a social partner than males with
633 shorter *avpr1a* alleles. Moreover, since the relationship between a male prairie vole's *avpr1a*
634 genotype and the number of females he sired offspring with was most evident among adult males
635 with the longest tenures (> 40 d) on the study site, long-lived males with longer *avpr1a* alleles
636 may have more social partners over their life (e.g., due to the death of a previous partner).

637 Our findings on the relationship between male *avpr1a* genotype and reproduction are in
638 contrast to those from the Solomon et al. (2009) study of semi-natural populations of prairie
639 voles, where parentage data indicated that males with shorter *avpr1a* microsatellite alleles sired
640 significantly more offspring with more females relative to males with longer *avpr1a* alleles. The
641 reasons for the discrepancy between these two studies of similar length are undetermined but
642 may result from the influence of length polymorphism at the *avpr1a* locus on male reproductive
643 behavior being very sensitive to ecological and other genetic variation. Densities within the
644 semi-natural enclosures were often much higher than in the Indiana population and Streatfeild et
645 al. (2011) showed that population density was one factor that significantly affected social
646 associations and mating patterns in prairie voles in Indiana and other natural populations.
647 Streatfeild et al. (2011) also detected correlations between vegetation characteristics and mating

648 patterns. The semi-natural populations were also exposed to less predation and migration out of
649 the population is prevented in these enclosed populations. Evidence suggests that sequence
650 differences in microsatellites of similar length (Phelps et al. 2010), as well as, genetic
651 polymorphism at other loci (Donaldson and Young 2013) influence the neural expression of
652 V1aR. The variation in these genetic factors between the lab-reared descendants of wild-caught
653 individuals from Illinois used in the Solomon et al. (2009) and the natural population of animals
654 in Indiana in this study is unknown. Unfortunately, partially due to the complexity found in
655 natural settings, we are unable to disentangle all the ecological and genetic variables that may
656 have affected sociosexual behavior between these two studies.

657 The greater number of offspring produced by males with longer *avpr1a* alleles suggest
658 that these males should have a higher fitness in natural populations and that, all else being equal,
659 the longer *avpr1a* alleles in males should be favored by selection. We did detect a skew in the
660 distribution of summed *avpr1a* lengths towards males with longer *avpr1a* alleles. The
661 distribution of *avpr1a* alleles in this population was statistically different from a normal
662 distribution, supporting the hypothesis that males with longer *avpr1a* alleles have a selective
663 advantage. However, without measures of *avpr1a* allele frequencies from this population across
664 many generations it is impossible to realistically assess how evolution is acting on allele size at
665 the *avpr1a* locus. Moreover, all things are not likely to be equal in nature. The greater
666 reproductive success by males with longer *avpr1a* alleles may be counterbalanced by other
667 fitness benefits associated with shorter *avpr1a* alleles in males or females. The greater
668 reproductive success of males with longer *avpr1a* alleles also may be conditional depending on
669 environmental factors, which may explain some of the discrepancies between this and the other

670 field studies examining the relationship between a male's *avpr1a* genotype and sociosexual
671 behavior.

672 Nonetheless, the data from our study does suggest that a male's *avpr1a* genotype is
673 associated with his reproductive success in nature under certain circumstances and we think this
674 is an important finding. While laboratory studies have provided a wealth of information on the
675 neurobiological mechanisms regulating social behavior, including behaviors associated with
676 social monogamy (e.g., Insel 1997; Young and Wang 2004; Young et al. 2005; Bales et al.
677 2007), the value of these studies to understanding the sociosexual behavior of prairie voles in
678 nature has been a matter of debate because it is unclear how these mechanisms regulate behavior
679 within the complexities of the natural world. The importance of the current findings are that they
680 are the first data to support the hypothesis that a male's *avpr1a* genotype is associated with
681 intraspecific variation in the genetic mating system and male reproductive success observed in
682 natural populations of prairie voles. The data from this study, in conjunction with the previous
683 studies that examined the relationship between male *avpr1a* genotype and sociosexual behavior
684 in the field, demonstrate the necessity of examining laboratory findings within ecologically
685 relevant contexts to determine if specific neurobiological mechanisms shown to regulate social
686 behavior in the laboratory are biologically relevant mediators of behavior in natural contexts.
687 Furthermore, these field studies highlight the potential difficulty in extrapolating findings on the
688 proximate basis of variation in sociosexual behavior from one natural population to another or
689 even from one time to another within a population.

690

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698

699 **Ethical standards** All trapping, handling and marking procedures were approved by the Miami
700 University Institutional Animal Care and Use Committee and were in accordance with the
701 guidelines of the American Society of Mammalogists for the use of wild mammals in research
702 (Sikes et al. 2011).

703 **Conflict of interest** The authors declare that they have no conflict of interest.

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705

706

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903

904 **Figure Captions**

905

906 **Fig. 1** Frequency distribution of summed length (bp) of an individual's two *avrpla*
907 microsatellite alleles for adult and subadult males ($N=168$) trapped in the study population

908

909 **Fig. 2** The probability that males in the study population with the shortest (■ = quartile 1) and
910 longest (■ = quartile 4) *avrpla* microsatellite allele lengths sired offspring with zero females,
911 one female or more than 1 female for each of the 4 tenure quartiles (1-4)

912

913 **Fig. 3** The probability that males in the study population with the shortest (■ = quartile 1) and
914 longest (■ = quartile 4) *avrpla* microsatellite allele lengths sired zero offspring, one offspring or
915 more than 1 offspring for each of the 4 tenure quartiles (1-4)

916

917

918

919 **Table 1** Results of linear (a) and logistic (b) regressions examining the effect of male *avpr1a*
 920 microsatellite allele length on field measures of adult male space use and social associations with
 921 adult females in free-ranging prairie voles

		<i>N</i>	<i>avpr1a</i> genotype
922			
923			
924			
925	Mean squared distance moved by		
926	males (a)	Period 1	40 $t=0.45; P=0.45$
927		Period 2	42 $t=-1.58; P=0.12$
928		Period 3	58 $t=0.71; P=0.48$
929			
930	Proportion of females in population		
931	overlapped by each male (a)	Period 1	32 $t=-0.03; P=0.98$
932		Period 2	39 $t=-0.10; P=0.92$
933		Period 3	50 $t=1.48; P=0.15$
934		Period 4	49 $t=0.58; P=0.56$
935			
936	Proportion of home range overlap		
937	between each male and the adult		
938	female he overlapped with the most (b)	Period 1	36 $z=1.20; P=0.22$
939		Period 2	44 $z=-0.23; P=0.82$
940		Period 3	55 $z=-1.72; P=0.08$
941		Period 4	49 $z=0.68; P=0.50$
942			
943	Number of females with which a		
944	male associated (b)	Period 1	13 $z=-1.22; P=0.22$
945		Period 2	25 $z=0.23; P=0.82$
946		Period 3	26 $z=1.80; P=0.07$
947		Period 4	34 $z=-0.68; P=0.50$
948			
949	Relative strength of a male's		
950	association with the most associated		
951	female (b)	Period 1	13 $z=1.21; P=0.22$
952		Period 2	25 $z=-0.23; P=0.82$
953		Period 3	26 $z=-1.72; P=0.84$
954		Period 4	34 $z=0.68; P=0.50$
955			
956	Male average residency score		
957	during all 4 trapping periods (b)		64 $z=-1.25; P=0.21$
958			
959			

960 **Table 2** Number of alleles (N_A), observed heterozygosity (H_o), expected heterozygosity (H_e) and
 961 polymorphic information content (PIC) among adult male and female prairie voles for the six
 962 microsatellite loci used in the genetic parentage analysis

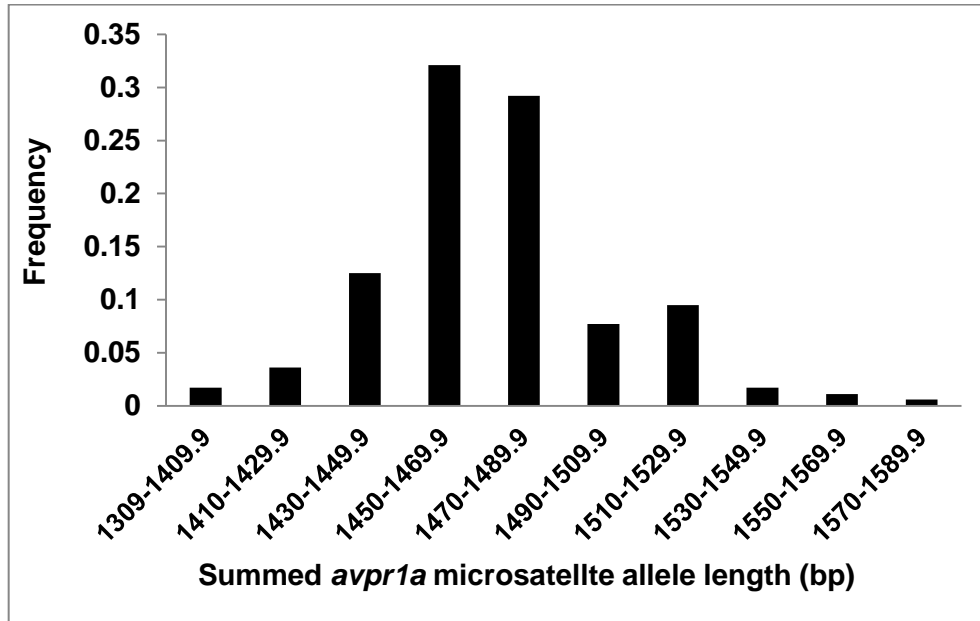
963	Locus	N_A	H_o	H_e	PIC
964					
965	AV13	27	0.887	0.896	0.887
966	MOE2	18	0.818	0.859	0.846
967	MSMM-2	24	0.796	0.905	0.896
968	MSMM-3	18	0.831	0.846	0.830
969	MSMM-5	18	0.743	0.910*	0.902
970	MSMM-6	28	0.265	0.428	0.428

971 * Locus deviates significantly from Hardy-Weinberg equilibrium (with Bonferroni correction)

972

973 Fig. 1

974



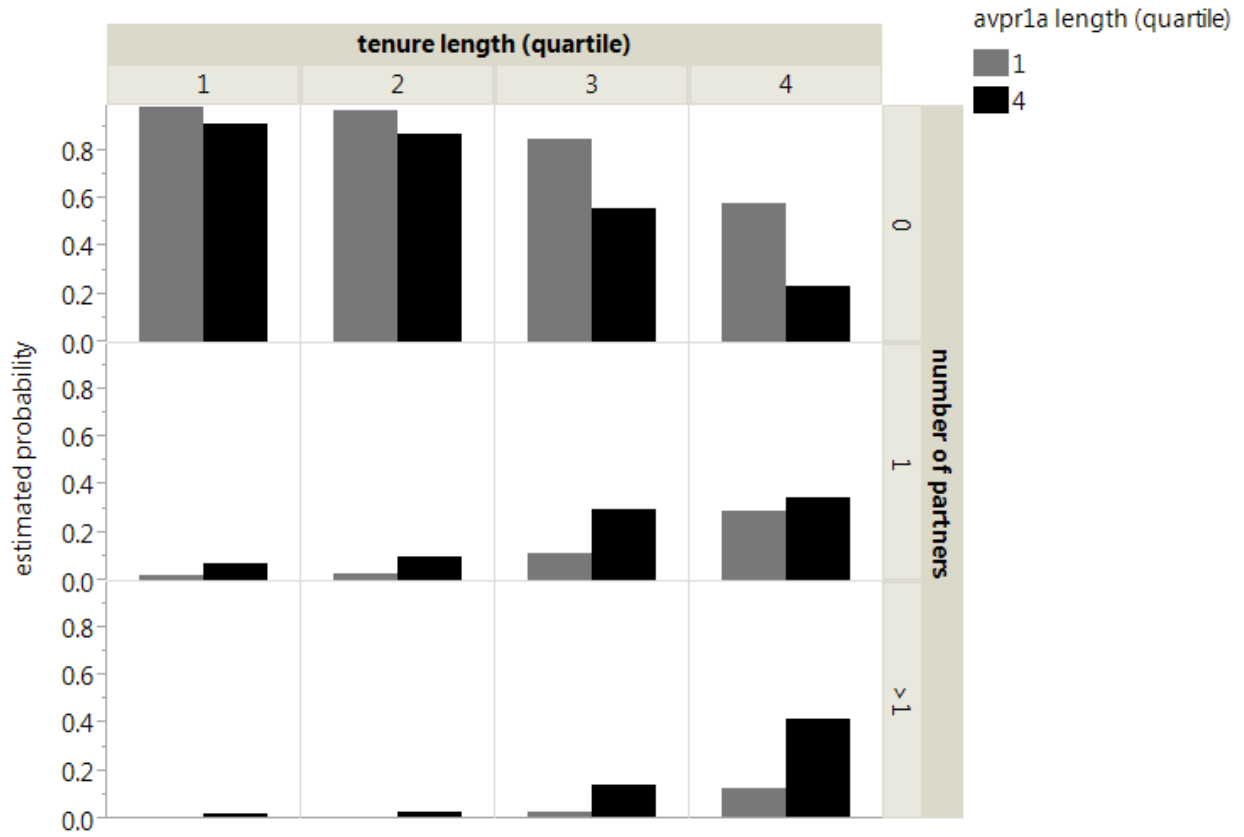
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978 Fig. 2

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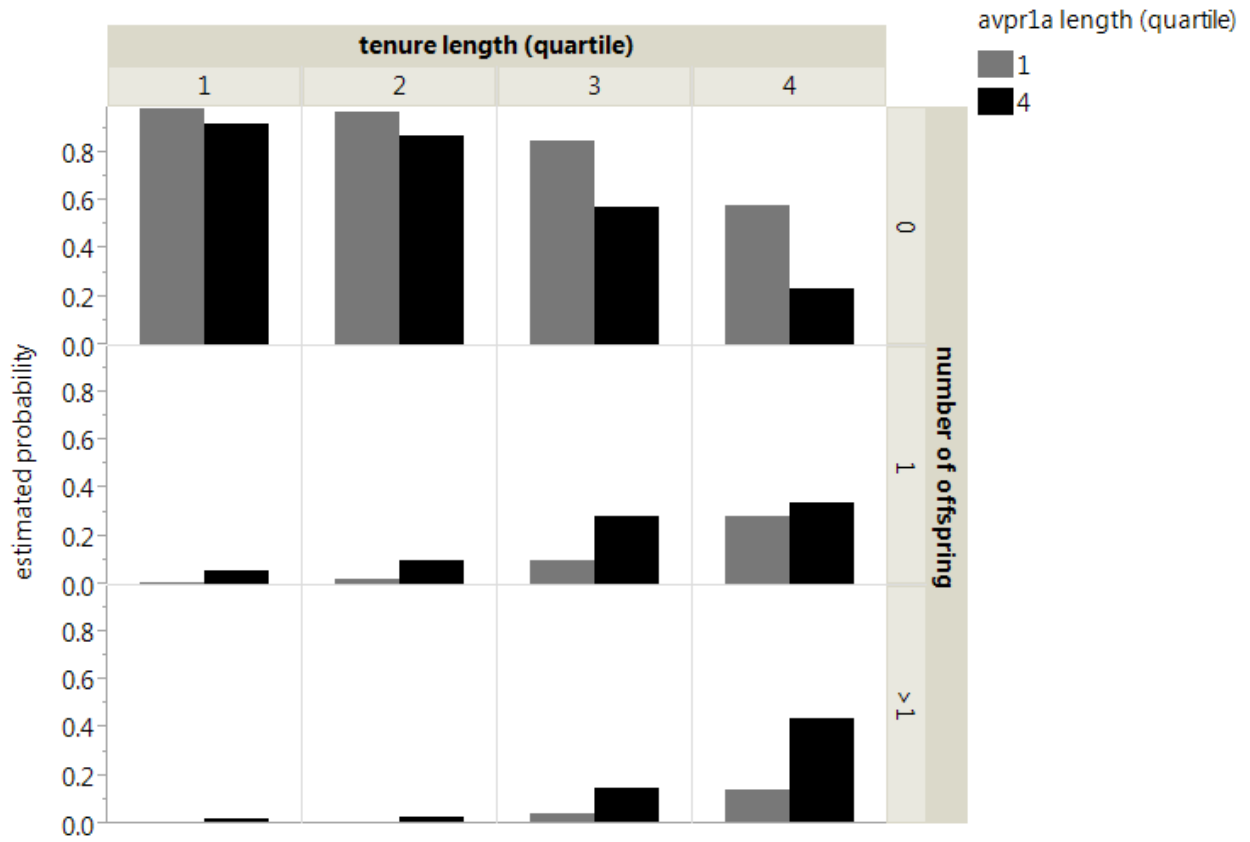
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985 Fig. 3



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